

**NUTRITIONAL VARIATIONS AMONG
DIFFERENT VARIETIES OF VELVET BEANS
(MUCUNA PRURIENS) WHEN EVALUATED AS
POTENTIAL PROTEIN SUPPLEMENTS
IN BROILER RATIONS.**

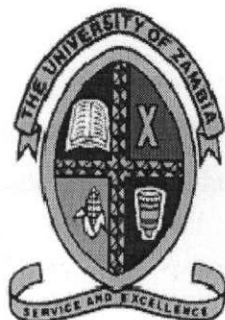
MUSONDA

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POTENTIAL PROTEIN SUPPLEMENTS IN BROILER RATIONS

BY

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A RESEARCH PROJECT REPORT SUBMITTED TO THE SCHOOL OF
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DEDICATION

This work is dedicated to the following people: My mother Annie Lombe, My husband Edwin Kikamba for being patient and encouraging, my daughter Patricia for understanding during my long absence from home.

ACKNOWLEDGEMENTS

I sincerely thank my supervisor, Dr. Simbaya for his input in this work.

My thanks go to Mpika boy's High School Administrators for their financial support. Staff of the Department of Animal Science at the University of Zambia also deserves special thank you for their contributions and positive criticisms which helped me complete this work.

Many thanks also go to Miss. Maliti for her technical assistance during laboratory experiments. I also extend my gratitude to my classmates for their support and guidance during the same period.

DECLARATION

This thesis has been compiled by myself and has not been accepted in any previous application for a degree at any university. I therefore, declare that this is a true reflection of my own work.

SIGNITURE..... *V. Musenah*

ABSTRACT

This study was carried out to evaluate nutritional differences among four different varieties of velvet beans (*mucuna pruriens*) i.e. yellow, speckled, cream white and black. 300g of each variety was ground to pass through a 2mm sieve and processed by soaking in 0.2% sodium carbonate for 24 hrs, then autoclaved at 120°C for 30 minutes to inactivate anti-nutritional factors. The varieties samples were evaluated for nutrient composition in triplicate using AOAC (1998). Proximate analysis results were as follows; dry matter ranged from 92.9% in yellow variety to 93.3% in black variety and these results showed no significant difference ($P < 0.05$). The results for other parameters showed significant differences ($P < 0.05$) and ranges were as follows; on CP the range was 21.77% in black to 29.08% in cream variety. On CF the range was from 8.61% in cream to 10.9% in black, Ash ranged from 2.77% in black to 3.21% in speckled and cream, EE ranged from 1.64% in cream to 4.00% in black, NFE ranged from 48.71% in cream to 53.09% in black variety and Metabolizable energy range was from 3.73kcal in black to 3.87 kcal in cream variety. The results for calcium ranged from 0.93% in cream to 1.32% in speckled while phosphorus ranged from 0.08% in speckled to 0.6% yellow. Calcium results did not show significance difference ($P > 0.05$) among treatments. Diets were then formulated by substituting soya at 20% at equal levels and a control diet containing soybean only was included, these were fed to Wistar rats for a period of 14days to determine DM and CP digestibility, feed intake, feed conversion ratio and change in weight. The results for DM apparent digestibility ranged from 83.0% yellow to 87.0% control diet, while CP apparent digestibility ranged from 74.0% in to 80.0% in control diet. The results showed no significant difference between diet containing speckled variety and the control diet. The average feed intake ranged from 14.28g in control diet to 18.50g in speckled diet per day, these results showed no significant difference ($P < 0.05$) between diet containing speckled and control diet. The mean change in weights ranged from 9.67g in diet containing speckled to 14.50g in speckled diet. These results showed no significant difference between speckled diet and the control diet. Apparent Feed conversion ratio (FCR) also ranged from 1.04 in control to 1.74 in black, the results showed no significant difference between speckled diet and control diet. This study demonstrates that the diet containing speckled variety had high apparent digestibility percentage in DM and CP, great increase in weight compared to soya diet and other diets containing other varieties of *mucuna pruriens*. Therefore, I recommend the speckled variety to be used in feed rations.

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ABBREVIATIONS AND ACRONYMS

CP.....	Crude protein
EE.....	Ether extracts
NFE.....	Nitrogen free extracts
CF.....	Crude fibre
FCR.....	Food conversion ratio
LSD.....	Least square difference
SEM.....	Standard error of the mean
ACF.....	Agriculture consultative forum
PAZ.....	Poultry association of Zambia
AOAC.....	Association of Analytical Chemists

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CHAPTER 1

1.0 INTRODUCTION

Broiler meat accounts for 90 % of Zambian poultry meat market (PAZ, 2010). However, intensive broiler production in Zambia has greatly been affected by high cost of production due to limited supply of feed ingredients, especially the conventional plant protein soya beans meal. The total annual production of soya beans in Zambia was 111,888 metric tonnes compared to 135,984 metric tonnes consumption for human and livestock in 2010/2011 (ACF, 2010). The total imports of soya beans were 29 metric tonnes, while the total import of stock feed with soya beans was 84 metric tonnes. The total national monthly consumption of soya beans was estimated at 11,332 metric tonnes in 2010, out of which 1,103 metric tonnes was for human consumption and 10,229 metric tonnes was for feed formulation (Agricultural Consultative Forum 2011). This shows that there is more soya beans being used in livestock feed rations than it is for human consumption and causing a deficit of 23,983 metric tonnes of required total consumption for both human and livestock. This has led to increased demand and high cost of soya beans.

The limited supply and high cost of soya beans as a conventional protein source in broiler diets has constituted a major economic concern to broiler chicken farmers. This has led to high cost of feed and hence increasing the total cost of production. The problem is further compounded by high level of adulteration of feed by some farmers in an attempt to supplement protein level, so that they can produce at a lower cost. For this reason, there is need to reduce on the amount of soya beans used in livestock rations by exploring alternative replacements.

The utilization of underutilized tropical legumes like velvet beans (*mucuna pruriens*) has been identified and used in poultry feed formulation because it possesses similar nutritional profile as soya beans (Tuleum *et al.*, 2008). However, there are different varieties of velvet beans that are grown in different parts of the world. In Zambia most varieties have been developed for soil fertility improvements such as sam (white seed coat), green and NIRS 16 (black). Many other wild varieties grow in different parts of Zambia on virgin lands (Nyirenda *et al.*, 2003). Thus, there is need to evaluate the nutritional differences among these different varieties of velvet beans on nutrient content, so that the best variety with acceptable nutrient levels could be used as protein source in feed rations to reduce on the quantity of soya beans used.

1.1 PROBLEM STATEMENT

The limited supply and high cost of soya beans as a conventional protein source in livestock diets has constituted a major economic concern to livestock farmers. This has led to high cost of feed and hence increasing the total cost of production. There is a need to look for alternative plant protein source if the cost of production is to be reduced.

1.2 JUSTIFICATION

Soya is in limited supply and often too expensive. Velvet beans identified as a potential alternative to soya beans, but its use is limited by low protein content and high levels of anti nutritional factors. Different varieties of velvet beans may have different levels of nutrients hence the need to look at the chemical composition and feeding quality characteristics of the various varieties of velvet beans. This will help to identify the best cultivar to be used in feed rations as plant protein source and reduce the use of expensive soya beans.

1.3 OVERALL OBJECTIVE

To evaluate the differences in the chemical composition, nutrient digestibility and feeding quality characteristics of different cultivars of velvet beans when used as alternative protein replacements to soya beans meal in broiler rations.

1.4 SPECIFIC OBJECTIVES

1. To determine the nutrient content found in different varieties of velvet beans.
2. To determine digestibility of nutrients in different varieties of velvet beans compared to soya beans.
3. To determine the feed intake, change in weight and feed conversion ratio for different varieties of velvet beans/varieties compared to soya beans.

1.5 HYPOTHESIS

- HO: There are no significant differences in the composition of nutrients among different varieties of velvet beans.
- HA: there are significant differences in the composition of nutrients among different varieties of velvet beans.

- HO: There is no significant difference in digestibility of nutrients found in different varieties of vello beans compared to soya beans.
- HA: There is a significant difference in digestibility of nutrients found in different varieties of vello beans compared to soya beans.
- HO: There is no significant difference in feeding quality characteristics of different varieties of vello beans compared to soya beans.
- HA: There is a significant difference in feeding quality characteristics of different varieties of vello beans compared to soya beans.

CHAPTER 2

2.0 LITERATURE REVIEW

Velvet beans (*mucuna pruriens*) are flowering legume plants and the flowers could be white, dark purple or light purple hanging in clusters. The plant produces clusters of pods containing seeds known as mucuna seeds. The name velvet bean is derived from a fact that the plant is covered in soft hairs when young. However, as the velvet bean matures, it loses these hairs. The leaves are ovate shaped, with sharp points and grooved sides.

2.1 Origin and Climatic conditions

Velvet beans originated from Southern Asia and it was introduced in southern states of United States of America in the late 19th century and in the tropics in the early 20th century (Elliotta *et al.*, 2003). Velvet bean is cultivated in tropical areas, such as the Caribbean, India and Africa. Velvet beans require warm temperature of 20°C to 30°C throughout the growing period, a frost free period of 180 to 240 days with an average rainfall of between 600 and 2500mm/year. A wide range of soil types are suitable, provided that they are well drained, since velvet beans cannot stand water logging. They tolerate fairly acid soils, with a pH of between 5 and 6.5 (Siddharya *et al.*, 1996).

2.2 Use of velvet beans

The crop is grown mixed with other vigorous growing crops such as Maize, to improve soil fertility through nitrogen fixation (buckles, 1995). In Zambia, it is grown to improve soil fertility prior to introduction of the chemical fertilizer-based Lima programme in Zambia (Kaonga, 2002). Velvet beans are nutritious animal feeds; the mature seeds are used in compound feeds after treating them to remove anti-nutritional factors, additionally the foliage can be fed to ruminants and non-ruminant animals (Eliita *et al.*, 2003, Chikangwa *et al.*, 2009). At this level, birds exhibit better growth performance in feed intake, weight gain, feed conversion ratio and protein efficiency ratio in both the starter and finisher diets (Vadivel *et al.*, 2011). Like any other beans, *M. pruriens* contains protein, vitamins and minerals making it an attractive and important source of plant protein for feeding animals (FAO, 1994).

2.3 Nutrient composition of velvet beans

The proximate composition of mature seeds contain; Moisture 10%, Protein 24.4%, Fat 5.7%, Nitrogen Free Extractives 5.5%, Fibre 6.4%, ash 3%, Calcium 0.18%, Phosphorus 0.99%, Potassium 1.3%, Vitamin A 50iu/100g, Thiamine 0.5mg/100g, Riboflavin 0.20mg/100g, Niacin 1.7mg/100g (Ravindran *et al.*, 1988) The oil present in the seeds have been found to be highly unsaturated with 47.2% linolic acid, 14.2% oleic acid, 3.8% Linolenic acid and 0.5% palmitoleic acid. The saturated fatty acids are Palmitic 19.5%, Stearic 12.6% and Arachidic 2.2%. Amino acids present are Isoleucine, leucine, lysine, Methionine, Cystine, Phenylalanine, Tyrosine, Threonine, Valine, Arginine, Histine, Alanine, Aspartic acid, Glutamic acid, Glycine, Proline and Serine. (Rehr *et al.* 1973)

2.4 Nutrient composition variation of velvet beans seeds

The composition of nutrients may vary slightly in different varieties of velvet beans especially in protein content. The difference is seen from proximate analysis of different varieties of *Mucuna pruriens* analysed by Dwight *et al.*, (2006) found mottled type to have 27.7% CP, while black and white was 25.8% CP. The analysis of nutrients in Zambian local varieties by Nyirenda *et al.*, (2003) also showed a difference in crude protein content were speckled had 24.95%, green 23.7% and black 22.5%. In the research by Tuleum *et al.* (2001), the crude protein in cream velvet beans was found to be 28.1% Cp in white and 24% Cp in speckled and black types.

2.5 Anti-nutritional factors of velvet beans

Velvet beans contain anti-nutritional factors like trypsin inhibitors, tannins and cyanide (Ravindran and Ravindran, 1988), anticoagulants (Houghton and Skari, 1984) and L- 3,4 dihydroxyphenylalanine (L Dopa) a potentially neurotoxin agent occurs in large amounts in mucuna (Carew *et al.*, 2003) Other anti-nutritional factors include haemagglutinin, chemotrypsin inhibitors, anti-vitamins, pepsinase inhibitors, phytic acid, flatulence factors and saponins (Emenalom and Janardhan, 2000). The anti-antinutritional factors phytate, trypsin and chemotrypsin adversely affect the protein digestibility; while L-Dopa causes vomiting and severe diarrhoea (Gupta, 1987). These substances unless destroyed by heat or some other suitable treatment can exert adverse physiological effects when ingested by animals (Liener 1980). However, it has been suggested that the consumption of velvet beans in low levels of certain anti-nutrients may produce health benefits while avoiding some of the adverse effects associated with their large intake (Thompson, 1988).

The feeding potential of the velvet can be enhanced by reducing these ant-nutritional factors using different treatment methods such as soaking in sodium hydroxide then autoclaving, boiling, and toasting (Olaboro *et al.*, 1991). According to Vadivel *et al.*, (2011), Soaking velvet beans in 0.2% sodium carbonate solution and exposing it to autoclaving treatment results in maximum reduction of various anti-nutritional substances.

CHAPTER 3

3.0 MATERIALS AND METHODS

3.1 LOCATION

The study was conducted at the University of Zambia in the School of Agricultural Sciences in Animal Science Department. The chemical composition analysis was done in the animal nutrition laboratory, and experiment for feeding trials with Wistar rats was also done in the same laboratory.

3.2 MATERIALS

Speckled, White, yellow and black velvet beans varieties were collected from Kasisi Agricultural Training Centre, while Soya beans cake, Dicalcium phosphate (DCP), methionine, broiler premix, salt, lysine, and limestone and cassava meal were procured from Livestock Services Co-operative Society in Lusaka. The Wistar rats were obtained from the Animal Care Unit of the Department Biological Sciences in the School of Natural Sciences at the University of Zambia.

3.3 PROCESSING OF VELVET BEANS

The processing of velvet beans was done by soaking 300g of velvet seeds of each variety in 0.2% sodium carbonate for 24 hrs after which the beans were dried and autoclaved at 120°C for 30 minutes using an autoclave in the Food Science Laboratory. The samples were ground to pass through a 2mm sieve.

3.4 CHEMICAL COMPOSITION ANALYSIS

Proximate analysis was done on processed velvet beans following AOAC methods (1998). All analyses were done in triplicates.

Moisture content was determined by drying 2g of each sample in a drying oven at a temperature of 105°C for 12 hours, the difference between the weight before and after drying was calculated to determine the moisture content, this was then expressed as a percentage of the original sample weight.

Protein content was determined using the kjeldahl method. Where 1g of each sample was digested in 12ml concentrated sulphuric Acid. The digest was then cooled to room temperature and 75ml distilled water was added and mixed thoroughly. Then 50ml of 40% sodium hydroxide was added to release ammonia. Volatilized ammonia was then collected in a receiving flask containing 25ml

of 4% boric acid indicator for 5 minutes to obtain ammonium borate. This was then titrated with 0.01N hydrochloric acid to the first trace of pink colour. To estimate the amount of Nitrogen the following formula was used;

Crude protein = (Titration - Blank) $\times$ 0.1 $\times$ 14.007 $\times$ 100 divide by sample weight. The Nitrogen content was converted to crude protein by multiplying by factor of 6.25.

Ether Extract was determined using soxhlet method. 2g (W1) of each sample was weighed and put in a pre-dried extraction thimble. The fat was extracted with 200ml of petroleum ether for 6 hours in a previously dried and weighed extraction flask (W2). After 6 hours, the solvent was evaporated. The flask was cooled and weighed (W3) and the crude fat was then calculated by applying the following formula; %fat = W3 - W2 divide by W1 $\times$ 100

Ash determination was done by combusting 2g in a muffle furnace for 4 hrs at 550°C. The difference in weight between the empty crucible and the crucible containing Ash gave the ash content of the sample.

Crude fibre was done by using Weende method, where by 0.5g of each sample was weighed (w) and placed in a beaker. 150 ml of 1.25% sulphuric acid was added and the contents in the beaker were heated over a hot plate. After boiling for 30 minutes, the contents were rinsed with 100mls hot water. The contents were washed back in the flask using 10mls sodium hydroxide and 150mls of 1.25% sodium hydroxide and the contents in the beaker brought back to boil again for another 30 minutes. The contents were then filtered through sintered crucibles, rinsed and the crucibles washed with 100mls of hot water. The crucibles were then dried in an oven overnight at 105°C (A). The crucible was then placed in a muffle furnace at 550°C for 1 hour to burn off remaining organic matter (B). The loss in weight on burning represented the fibre in the original sample. The crude fibre was then expressed as percentage by mass using the following formula; % crude fibre = (A - B)/W $\times$ 100. Gross energy was obtained by summation of the known energy contributed by the analysis of crude protein, crude lipid and carbohydrate fractions of the sample. This was calculated as follows;

Energy contributed by crude protein = protein content sample % $\times$ 5.4 kcal/g = X

Energy contributed by crude lipid = lipid content of sample % $\times$ 9.5 kcal/g = Y

Energy contributed by carbohydrate was calculated as nitrogen free extract using the following formula;

NFE (crude carbohydrate) = $100 - (\% \text{ moisture} + \% \text{ crude protein} + \% \text{ crude lipid} + \% \text{ Ash})$.

Then energy contributed by crude carbohydrate = energy in crude protein sample $\times 4.1 \text{ kcal/g} = Z$

The energy values given for each component (kcal/g) are the average values obtained for a whole range of types of material when 1g is completely combusted in a calorimeter. Gross energy (GE kcal/g) = $(X + Y + Z)/100$.

Calcium and Phosphorus were determined by weighing 1g ash sample, added 10mls of 3N HCl and boiled to yellow colour. The sample was filtered, 5mls was taken and 100mls of distilled water was added to fill up to the mark. 1g of ammonium Oxalate, drops of methyl red then boil, cool and filter. At this point calcium was determined by adding 25mls ammonium sulphate and titrated to pink colour while solution was boiling using potassium permanganate. Phosphorus was determined using Vanado-molybdate method spectrophotometrically. The chemical composition analysis is shown in appendix 1.

3.5VELVET BEANS DIGESTIBILITY AND FEEDING TRIAL

3.5.1EXPERIMENTAL DESIGN

30 Wistar rats were randomly allocated to the 5 dietary treatments with 6 replications per treatment in a completely randomised design (CRD) and each rat was treated as experimental unit on which data was collected.

3.5.2 MANAGING WISTAR RATS

Feed and water was offered Adlibitum. The rats were subjected to standard management procedures and each rat was taken as individual replication. The feeding trial lasted for a period of 10 days during which feed intake was recorded daily. Feed intake was determined by subtracting the left over feed on the following day from the quantity (30g) given on the previous day. Faecal matter was collected daily dried and weight recorded. The tables for average feed intake faecal matter recorded are shown in the appendix 2.0 and 3.0 respectively.

The initial body weight of the experimental rats was taken prior to the commencement of the experiment, and the final body weight gain was determined at the end of the experiment after 10 days. Changes in body weight were determined by subtracting initial body weight from final body weight. The results obtained are also shown in the appendix 5.0.

The apparent feed conversion ratio was then calculated from data collected for feed intake and data collected from change in weight for a period of 10 days. Feed conversion ratio was then determined by dividing the average feed intake by average body weight gain.

3.5.3 DIET COMPOSITION

The four treatment diets contained different varieties of velvet beans at the same inclusion level to replace soya beans at 20% and fifth treatment diet of soya beans only was included as a control. Velvet beans and soya bean meal contributed 45%, while basal diet contributed 55% in all the treatments. The dietary composition and the calculated analysis of nutrient content are shown in the following tables.

Table 1.0 Dietary Composition (%) of different treatment diets.

INGREDIETS	TREATMENT DIETS				
	YELLOW	SPECKLED	CREAM	BLACK	SOYA
Soya bean cake	25	25	25	25	45
Velvet meal	20	20	20	20	—
Cassava meal	42.1	42.1	42.1	42.1	42.1
Fish meal	5	5	5	5	5
Soya oil	5	5	5	5	5
limestone	0.58	0.58	0.58	0.58	0.58
Dicalcium phosphate	1.82	1.82	1.82	1.82	1.82
Lysine	0.08	0.08	0.08	0.08	0.08
Methionine	0.08	0.08	0.08	0.08	0.08
Broiler premix	0.16	0.16	0.16	0.16	0.16
Salt	0.16	0.16	0.16	0.16	0.16
total	100	100	100	100	100

Table 2.0 calculated analysis of nutrient content in diet.

Energy	3,089 Kcal
Crude protein	19%
Calcium	18.2g
phosphorus	5.8g
Methionine	0.8g
Lysine	0.8g

3.5.4 APPARENT DIGESTIBILITY OF NUTRIENTS.

Proximate analysis in diet was done and the results were used in calculating digestibility of nutrients in the treatments.

Apparent digestibility was calculated by subtracting amount of nutrient in faecal output from the amount of nutrients in feed intake, divided by feed intake and then converted to a percentage McDonald *et al.*, (1986).

3.6 DATA ANALYSIS

Analysis of variance (ANOVA) was used in the analysis of quantitative data using the Genstat 14th edition. For multiple comparison and differentiating means with and among treatments the least square difference (LSI) was used and Duncan's multiple range test was used to separate statistically different means.

CHAPTER 4 RESULTS AND DISCUSSION

4.1 CHEMICAL COMPOSITION ANALYSIS

The results for chemical composition of velvet beans are shown in following table.

Table 2.0 chemical composition (%) mean + SEM).

Parameter	varieties				
	Yellow	Speckled	Cream	Black	SEM(+/-)
DM	92.9	93.2	93.1	93.3	± 0.15
Ash	3.13 ^b	3.21 ^b	3.21 ^b	2.77 ^{ab}	± 0.10
Cp	24.98 ^b	24.66 ^b	29.08 ^c	21.77 ^a	±0.29
EE	3.29 ^b	3.09 ^b	2.64 ^a	4.00 ^c	±0.13
CF	10.73 ^b	9.07 ^a	8.61 ^a	10.99 ^b	±0.39
NFE	50.77 ^b	53.57 ^c	48.71 ^a	53.09 ^c	±0.56
ME/kcal	3.75 ^a	3.83 ^b	3.87 ^b	3.73 ^a	±0.04
Calcium	1.00 ^a	1.32 ^b	0.93 ^a	1.17 ^{ab}	±0.07
Phosphorus	0.61 ^d	0.08 ^b	0.48 ^c	0.48 ^c	±0.01

Note. Values with different superscript within rows are significantly different ($P < 0.05$).

The results for all the nutrients analysed showed a significant difference ($P < 0.05$) except in moisture content. The data was further subjected to least square difference (LSD) for multiple comparisons and Duncan Multiple range for separation of means.

Crude protein results showed no significance difference ($P > 0.05$) between yellow and speckled variety while black and white differed significantly ($P < 0.05$) from each and from two other varieties. Black velvet recorded lowest protein content of 21.77% (black), and 29.97% (cream) which recorded the highest.

The ether extract values showed no significant difference ($P > 0.05$) between yellow and speckled while black and cream differed significantly ($P < 0.05$) from each other and from the two other varieties. The lowest was cream 2.63% and highest was black with 4.00%.

The Crude fibre results showed no significance difference ($P > 0.05$) between yellow and black but there was a significant difference ($P < 0.05$) between the subset groups. The highest value was recorded in black with 10.91% and lowest was cream with 9.06%.

Ash content among the varieties showed no significant difference ($P > 0.05$) among three varieties yellow; speckled and cream while black differed significantly ($P < 0.05$) from the rest and recorded the lowest value of 2.77%.

Results on Nitrogen free extractives (NFE) results showed that, yellow and cream were found to be significantly different ($P < 0.05$) from each other and from other varieties while speckled and black did not differ significantly ($p < 0.05$) from each other. The lowest value was obtained as 48.71% cream and the highest was 53.57% in speckled.

Results on Metabolizable energy results showed that yellow and black did not differ significantly ($P > 0.05$) and also speckled and cream did not differ but there was a significant difference ($P > 0.05$) between the two groups.

Calcium results showed that there was no significant difference ($P > 0.05$) among yellow, speckled and black. But speckled differed significantly from the rest with 1.32%.

The results for phosphorus showed the least value of 0.08% in speckled variety and this differed significantly from the rest.

4.2 APPARENT DIGESTIBILITY OF NUTRIENTS RESULTS

Table 3.0 Apparent digestibility (%) for protein and dry matter, mean + SEM.

Treatment	Dry matter	Crude protein
Yellow	85.1 ^a ± 0.09	76. ^{ab} ± 0.317
Speckled	83.0 ^a ± 0.09	74.0 ^a ± 0.317
Cream	84.1 ^a ± 0.09	75. ^{ab} ± 0.317
Black	83.2 ^a ± 0.09	75.1 ^{ab} ± 0.317
soya	87.0 ^c ± 0.09	80.0 ^c ± 0.317

Note. Values with different superscripts within columns are significantly different ($P < 0.05$).

Apparent digestibility results showed no significant differences ($P > 0.05$) for crude protein dry matter among the different varieties of velvet beans, but control diet differed significantly from the rest.

4.3 FEEDING AND GROWTH PERFORMANCE

Table 4.0 shows the results obtained for feed intake, weight gain and Food Conversion Ratio (FCR) Mean+ SEM.

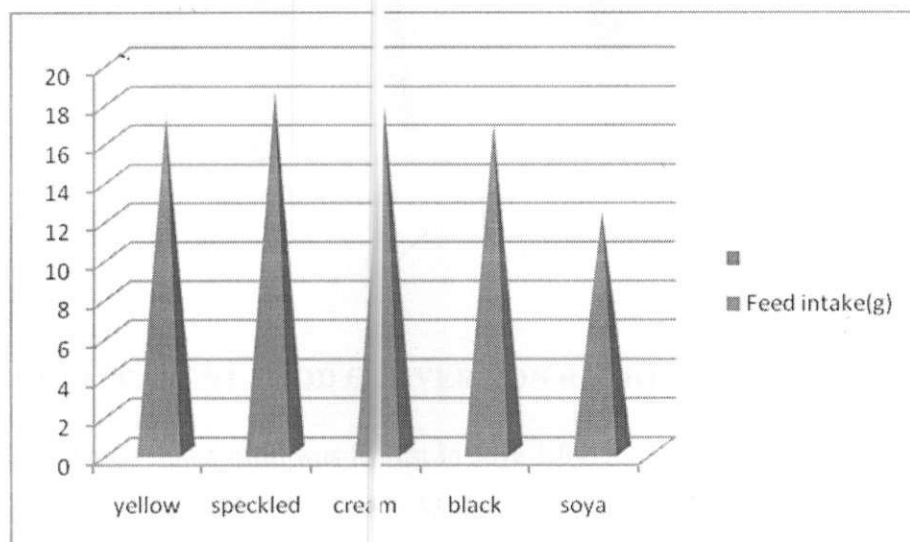
Parameter		treatment (variety of velvet + soya)				
	Yellow	speckled	cream	black	soya	SEM
Feed intake (g)	17.19 ^b	18.50 ^b	17.63 ^b	16.79 ^{ab}	14.28 ^a	± 0.932
Weight gain (g)	11.50 ^b	14.50 ^a	10.17 ^{bc}	9.67 ^{bc}	13.67 ^a	± 2.204
FCR	1.49 ^{ab}	1.28 ^{ab}	1.73 ^b	1.74 ^b	1.04 ^a	± 0.256

Note. Values with different superscript within rows are significantly different ($P < 0.05$).

4.3.1 FEED INTAKE

The results for feed intake showed no significant differences ($P > 0.05$) of the three treatment diets containing yellow, cream and black varieties, while control diet differed significantly ($P < 0.05$) from other treatments by recording the lowest ratio of 14.28g and the highest was recorded in speckled with 18.50g.

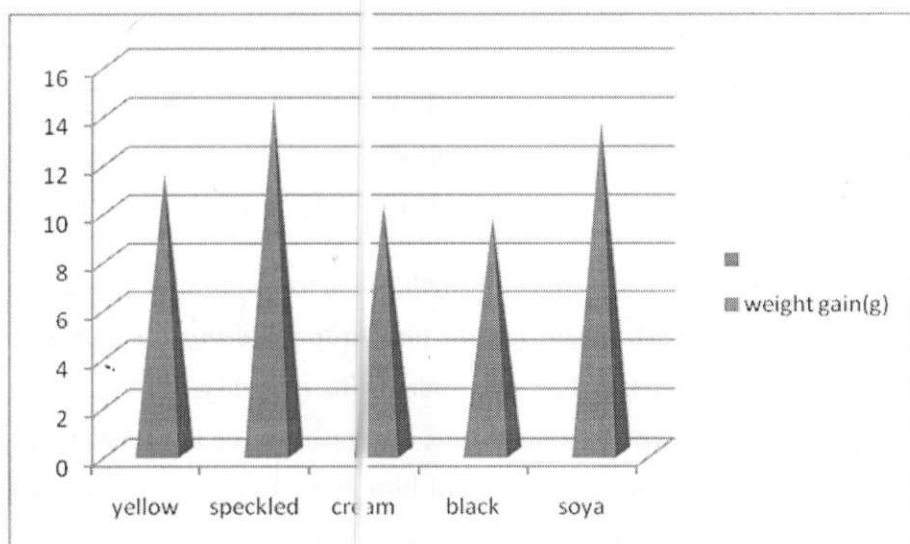
Figure 2.0 Average feed intake (g) per day.



4.3.2 WEIGHT GAIN

The highest weight gain of 14.5g was recorded in speckled and this was followed by control diet with 13.67g. This could be as a result of high feed intake, and low feed conversion ratio in speckled diet. While weight gain in control diet could be as a result of lowest FCR and highest protein digestibility recorded.

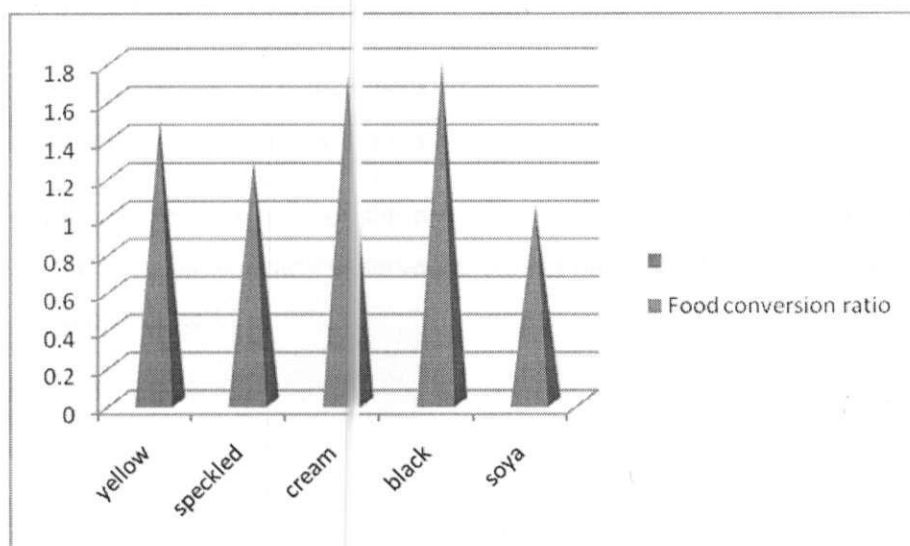
Figure 4.0 weight gain (g) for period of 10 days.



4.3.3 APPARENT FOOD CONVERSION RATIO

Food conversion ratio was lowest in soya 1.04, followed by diet containing speckled velvet bean meal with 1.28. Low FCR in speckled may as a result of high feed intake and high weight gain.

Figure 3.0 Apparent Food conversion ratios.



CHAPTER 5 CONCLUSION AND RECOMMENDATION

5.1 CONCLUSION

The research demonstrated that, there is a significance difference among the varieties of velvet beans (*mucuna pruriens*) in contents of nutrients. But this difference is not directly related to growth performance when used in feed rations.

The use of different varieties in diets produced results that were not significantly different in feed in apparent digestibility of protein and dry matter. However, the results for food conversion ratio (FCR) and weight gain of Control diet and diet containing speckled velvet meal did differ significantly when subjected to least square difference. The highest protein digestibility was recorded in control diet followed by speckled. Average feed intake was highest in speckled diet and lowest in control diet. Feed conversion ratio was lowest in control diet followed by speckled and weight gain was highest in speckled diet followed control diet. The results shows that speckled velvet beans can be used to replace soya beans in feed ration because of its low feed conversion ratio and high weight gain recorded.

Therefore, the conclusion is that speckled velvet beans can produce better results in growth performance if included in diets at 20% among other varieties, though other varieties can still be used were speckled variety is not available.

5.2 RECOMMENDATIONS

The recommendation is that speckled variety can be included in the diets to replace soya beans at 20% so as to reduce on the amount of soya bean to use in feed formulations.

I also recommend that, a similar research be conducted in day old chicks (broilers) for a period of six weeks so as to clearly observe the growth rate of birds, unlike the way it was used on Winstar rats for 10 days.

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7.0 APPENDICES

APPENDIX 1.0 Chemical composition in velvet beans

Parameter	Treatment 1 (yellow)			Treatment 2 (speckled)			Treatment 3 (cream)			Treatment 4 (black)		
	Replication			Replication			Replication			Replication		
	1	2	3	1	2	3	1	2	3	1	2	3
Moisture.	7.19	7.10	6.90	6.40	6.43	6.40	6.90	6.74	6.99	6.94	7.15	7.03
Ash	3.00	3.25	3.14	3.34	3.20	3.10	3.30	3.24	3.10	2.75	2.90	2.65
Cp	25.39	24.51	25.04	24.77	23.99	25.21	29.41	29.94	30.55	21.36	22.24	21.71
EE	3.40	3.18	3.30	3.20	2.76	3.30	2.84	2.34	2.72	4.18	3.84	3.98
CF	10.99	11.20	10.00	8.99	9.60	8.60	7.80	8.60	9.40	10.40	11.80	10.77
NFE	50.03	50.75	51.53	53.30	54.02	53.39	49.75	49.14	47.25	54.37	52.07	52.83
kcal/kg	3.75	3.71	3.73	3.83	3.77	3.89	3.90	3.85	3.85	3.78	3.70	3.72
Ca	1.04	0.96	1.00	1.46	1.12	1.38	0.84	0.99	0.96	1.10	1.06	1.34
P	1.01	1.1	1.12	0.40	0.38	0.39	0.54	0.51	0.55	0.46	0.44	0.48

APPENDIX 2.0 chemical compositions in diet (%).

Treatment	Crude protein	Crude fibre	Ether Extract	NFE	Ash
Yellow	18.95	4.70	10.28	51.54	5.90
Speckled	18.98	4.00	13.16	48.53	6.50
White	18.65	2.54	11.16	52.87	6.10
Black	18.56	4.20	11.24	51.60	5.80
Soya	18.91	2.30	12.2	49.81	7.95

APPENDIX 3.0 chemical compositions in faecal matter (%).

Treatment	Crude protein	Crude fibre	Ether Extract	NFE	Ash
Yellow	30.38	11.2	2.60	25.92	18.0
Speckled	29.50	8.10	2.54	30.36	18.1
White	28.19	7.80	1.98	32.85	17.38
Black	27.66	10.30	2.08	30.52	18.14
Soya	25.74	7.55	2.64	31.32	22.00

APPENDIX 4.0 Average feed intake (g) per day.

Treatment	Replications					
	1	2	3	4	5	6
Yellow	18.05	15.36	15.96	14.27	20.17	19.36
Speckled	19.39	16.02	19.10	14.63	22.95	18.93
White	18.82	18.26	19.44	15.63	15.20	18.42
Black	14.79	15.17	20.03	17.26	15.70	17.78
soya	12.69	12.10	18.11	14.00	13.14	15.66

APPENDIX 5.0 Change in weight gain (g) for a period of 10days.

Treatment	Replication					
	1	2	3	4	5	6
Yellow	19.0	9.0	12.0	9.0	7.0	15.0
Speckled	11.0	21.0	28.0	8.0	8.0	11.0
White	11.0	8.0	20.0	9.0	6.0	7.0
Black	6.0	14.0	7.0	11.0	7.0	13.0
Soya	12.0	13.0	14.0	14.0	8.0	21.0